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# EVALUATION OF ANTIBACTERIAL ACTIVITY OF LEMON JUICE ON SOME CLINICAL ISOLATES

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## ABSTRACT

This study was carried out to evaluate the antibacterial effectiveness of lemon juice, (ripe and unripe) extracts; using agar disc diffusion method. The test organisms (*Pseudomonas aeruginosa, Staphylococcus aureus, Escherichia coli, Bacillis subtilis* were obtained from National Agency for Food, Drug Administration and Cotrol Agulu, Anambra State. A serially diluted 24 hours broth culture of the test organisms was used to inoculate Mullar Hinton agar; and 10mm diameter paper disc impregnated with test organisms lace in it before incubation. The result of the test showed that the ripe lemon juice inhibited *Staphylococcus aureus* (30mm), *Pseudomonas aeruginosa* (28mm), *Bacillus subtilis* (20mm) but no inhibition on *Escherichia coli* while unripe lemon juice inhibited *Staphylococcus aureus* (24mm), *Pseudomonas aeruginosa*. (30mm), *Escherichia coli* (23) but did not inhibit *Bacillus subtilis*. The MIC and MBC carried out using ripe lemon on *Staphylococcus aureus* gave 30%v/v for both; while unripe lemon recorded 30%v/v and 15%v/ MIC and MBC results respectively on *Pseudomonas aeruginosa*. The result of this work has shown that lemon juice (both ripe and unripe) can be used in the treatment of infections caused by these organisms. However, more specific researches are needed for full application in the pharmaceutical industry.

Key Words: Lemon, Antibacterial, Inhibition.

## INTRODUCTION

For a long period in history, plants have been valuable and indispensable sources of natural products for the health of human beings and they have a great potential for producing new drugs (Nascimento *et al.*, 2000). Bacteria have the genetic ability to transmit and acquire resistance to drugs, which are utilized as therapeutic agents (Abeysinghe, 2010). Finding new naturally active components from plants or plant-based agricultural products has been of interest to many researchers. Hence, a great deal of attraction has been paid to the antibacterial activity of citrus as a potential and promising source of pharmaceutical agents (Ortuño *et al.*, 2006).

According to World Health Organization, medicinal plants would be the best source to obtain a variety of drugs. About 80% of individuals

from developed countries use traditional medicine, which has compounds derived from medicinal plants. Therefore, such plants should be

investigated to better understand their properties, safety and efficiency (Nascimento et al., 2000).

Lemon is an important medicinal plant of the family Rutaceae. It is used mainly for its alkaloids, which are having anticancer activities and

the antibacterial potential in crude extracts of different parts (leaves, stem, root, juice, peel and flower) of Lemon against clinically

significant bacterial strains has been reported (Kawaii et al., 2000). Citrus flavonoids have a broad spectrum of biological activity including

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antibacterial, antifungal, anti-diabetic, anticancer and antiviral activities (Burt, 2004).

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Antimicrobial activity of the peel extract is directly concerned with the components that they contain. The studies showed that essential oils, protopine and corydaline alkaloids, lactons, polyacetylene, acyclic sesquiterpenes, hypericin and pseudohypericin compounds are effective toward various bacteria (Keles *et al.*, 2001). Furthermore, citrus fruit had been used in traditional Asian medicines for centuries to treat indigestion and to improve bronchial and asthmatic conditions (Kalpa *et al.*, 2012).

Johann *et al.* (2007) have shown that citrus varieties are considered and containing a rich source of secondary metabolites with the ability to produce a broad spectrum of biological activities.

Extracts of citrus fruit (e.g. lemon, orange and grape fruit) are among the most studied natural antimicrobials for food app

### Statement of the problem

Recently, microbes have the ability of resisting the effects of medication, because the roles of medicinal plants have been neglected and despite the leading role of medicinal value of lemon and lime juice in pharmaceutical industries, less than 3% have been incorporated into drug manufacture, because this plant contain some bioactive compound that can serve as antibacterial agents, which can be used to prevent and treat bacterial infections, and antibiotics resistance occurs when bacteria change in response to the use of these medicines, therefore this deemed it necessary to compare the antimicrobial activities of lemon juice on some clinical isolates.

## **Objectives of the study**

This study achieved the following specific objectives:

- i. To extract juice from lemon
- ii. To assess their antimicrobial activities on some clinical isolates.

## LITERATURE REVIEW

## An Overview of Herbal Medicine

World Health Organization define Traditional herbal medicines as naturally occurring, plant-derived substances with minimal or no industrial processing that have been used to treat illness within local or regional healing practices. Traditional herbal medicine and their preparations 2
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have been widely used for the thousands of years in developing and developed countries owing to its natural origin and lesser side effects (Tilburt *et al.*, 2008). These medicines initially took the form of crude drugs such as tinctures, teas, poultices, powders, and other herbal formulations. The use of plants for healing purposes predates human history and forms the origin of much modern medicine. Clinical, pharmacological, and chemical studies of these traditional medicines, which were derived predominantly from plants, were the basis of most early medicines such as aspirin (willow bark), digitoxin (from foxglove), morphine (from the opium poppy), quinine (from cinchona bark), and pilocarpine (Jaborandi). Herbal medicine is still the mainstay of about 75 - 80% of the world population, mainly in the developing countries, for primary health care. This is primarily because of the general belief that herbal drugs are without any side effects besides being cheap and locally available. According to the World Health Organization (WHD), the use of herbal remedies throughout the world exceeds that of the conventional drugs by two to three times (Pal *et al.*, 2003).

Indigenous herbal medicines are those which historically used in a local community or region and are very well known through long usage by the local population in terms of its composition, treatment and dosage. It can be used freely by the local community or in the local region. However, if the medicines in this category enter the market or go away from the local community or region, they have to meet the requirements of safety and efficacy as per the national regulations for herbal medicines. Herbal medicines in systems have been used for a long time and are documented with their special theories and concepts, and accepted by the countries. Modified herbal medicines have been modified in shape, or form including dose, dosage form, mode of administration, herbal medicinal ingredients, methods of preparation and medical indications. They have to meet the national regulatory requirements of safety and efficacy of herbal medicines. Imported products with herbal medicine base covers all imported herbal medicines including raw materials and products (Pal *et al.*, 2003).

Plants and natural products were used by humankind over the years as food and medicines to cure and prevent diseases. It is very difficult to point out an exact time when the use of plants was started as medicine, the Carbon dating from ancient Babylon (Iraq) records that plants were cultivated as medicines 60,000 years ago. Written material medical of medicinal herbs go back approximately 5,000 years in India, China and Egypt and at least 2,500 years in Greece and Asia Minor. Neanderthal remains have been found to contain the remnants of medicinal herbs. Sumerians described well-established medicinal uses for plants such as laurel, caraway, and thyme at least 5,000 years ago. Egyptian people where well known to medicine before 2900 BC, these people used papyrus (pithy stem of a water plant for writing or FEDERAL POLYTECHNIC OKO, ANAMBRA STATE

painting) such as the Ebers papyrus(1500 BC), the Edwin Smith papyrus (1600 BC), the Berlin papyrus (1200 BC), and the Kahun papyrus (1900 BC), etc but the best known Egyptian pharmaceutical record is the "Ebers Papyrus" contain more than 800 formulae such as gargles, snuffs, poultices, infusions, pills and ointments, with beer, milk, wine and honey being commonly used as vehicles and 700 different drugs like acacia, castor oil and fennel etc with their uses along with apparent reference to some chemical such as iron, sodium chloride and sulphur.

Currently more than 80% of the world population depends on traditional and plant derived medicine because. Plants are important sources of medicines and presently about 25% of pharmaceutical prescriptions in the United States contain at least one plant-derived ingredient. In the last century, roughly 121 pharmaceutical products were formulated based on the traditional knowledge obtained from various sources. In fact, it is now believed that Nature contributes up to 90% to the new drug molecule. Nature has provided many of the effective agent such as dactinomycin, bleomycin, and doxorubicin, vinblastine, irinotecan, topotecan, etoposide, and paclitaxel (anticancer), Mefloquine chloroquine , amodiaquine artemisinin, dihydro artemisinin, artemether, and arteether (antimalarial) , metformin and eventually the other Biguanide, Harunganin, cryptolepine, maprouneacin (anti diabetic) Calanolide A, cucrcumin, phenethyle isocyanate, phenoxidiol (anti-HIV drugs) etc.

#### Importance of herbal medicine

Many herbal medicines have been used for hundreds of years and it is assumed in many cases that they must work. For example, about 7000 species of plants are used in China as herbal medicines, but only 230 of the most commonly used ones have been subject to in-depth pharmacological, analytical and clinical studies.

The consumption of herbal medicines is increasing steadily throughout the world as an alternative treatment for alleviating a number of health problems including heart diseases, diabetes, high blood pressure and even certain types of cancer. In India use of herbal drugs is much more because of their easy accessibility. Unlike drugs, herbal products are not regulated for purity and potency. There are neither studies on their effectiveness nor control over the quality and safety of these preparations. As per Food and Drug Administration mandates, only medicines have to be proven to be safe before being released into market. Herbal products do not fall under the category of medicine as long as they are not marketed for the prevention of any disease. Herbal drugs are considered as 'food integrators and readily available

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in the market without prescription. The major driving force for the use of herbal drugs is the perception that 'they are safe because they are natural and have fewer side effects than prescription drugs'. However, various studies and researchers have high lightened their possible side effects, if taken irregularly, in excessive amounts or in combination with some medicines (Stickel *et al.*, 2005).

A common problem with herb use is that people do not take into consideration how they may interact with any prescription drug they are taking, or with each other. Interaction between drugs and herbs can result in unexpected concentration of drugs and also cause undesired effects. Sometimes the use of commonly used herbs with prescription medicines become big barrier for the diagnosis of certain diseases as people do not inform their physicians about their consumption. The aim of this paper is to highlight the uses and side effects of some selected below mentioned herbal drugs so that these may be used safely (Stickel *et al.*, 2005).

#### Distribution and taxonomy of Citrus species

According to statistics of FADSTAT, Citrus species are grown all over the world in more than 140 countries, with more than 8.7 million hectares and about 131 million tons of fruits produced in 2012. And China, Brazil, the U.S.A., India, Mexico, and Spain are the world's leading Citrus fruit-producing countries, representing close to two-thirds of global production. In China, citriculture has existed traditionally, and the Citrus varieties have been naturally selected: (1) *C. aurantifolia* (Christm.) Swingle, (2) *C. aurantium* L., (3) *C. hongheensis* Ye *et al.*, (4) *C. hystrix*OC., (5) *C. ichangensis*Swingle, (6) *C. junos*Sieb. ex Tanaka, (7) *C. limon* (L.) Burm. f., (8) *C. limonia*Osb., (9) *C. macroptera*Montrous., (10) *C. maxima* (Burm.) Merr., (11) C. medica L., (12) *C. paradis*Macf., (13) *C. reticulata* Blanco, (14) *C. sinensis* (L.) Osb (Caristi *et al.*, 2003).

The genus Citrus belongs to the subtribeCitrinae, tribe Citreae, subfamily Aurantioideae of the family Rutaceae. However, continual taxonomic study appears to be very complicated and controversial, mainly due to sexual compatibility between Citrus species and related genera, the high frequency of bud mutations, apomixis (e.g., adventitious embryony). Consequently, there has been no consensus among taxonomists as to the actual number of Citrus species. Later, phylogenetic analysis indicated only three true species within the cultivated Citrus, i.e., *C. medica* L. (citron), C. reticulata Blanco (mandarin) and C. maxima (Burm.) Merr. (pummelo). In order to be convenient, the existing taxonomic systems are combined currently. Because morphological characters are of limited use, studies have mainly focused on new taxonomy methods, i.e., chemotaxonomy. 66 Citrus species and near-Citrus relatives can be cited in accordance with Tanaka's classification system with 24 flavonoids. Flavanones were used as chemotaxonomic markers to distinguish 77 Zhishi (traditional Chinese

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medicine) samples from three Citrus species. Another study suggested that the content of certain monoterpenes could be as taxonomic markers between *C. sinensis* and *C. junos*(Liu *et al.*, 2014).

#### Active secondary metabolites

Plentiful active natural metabolites including flavonoids, alkaloids, coumarins, limonoids, carotenoids, phenolic acids and essential oils, have been found in Citrus fruits. Tables in additional files have summarized these secondary metabolites isolated from peel, pulp, seed, pressed oil, juice or whole fruit from 31 common species to give a systematical profile. By these at least, the types of Citrus-derived secondary metabolites vary among different Citrus species and different fruit parts. Moreover, flavanones, synephrine, auraptene and limonin are the most dominants among the flavonoids, alkaloids, coumarins and limonoids groups, respectively.

In Additional file 1, 48 types of flavonoids from 22 common Citrus species of different fruit parts (peel, pulp, seed, pressed oil, juice or whole fruit) have been summarized. These flavonoids belong to the five classes: flavones, flavonols, flavonones, flavonones and polymethoxylated flavones. Anthocyanins, an uncommon class of flavonoid, only appears in blood oranges of limited data in different fruit parts. Among Citrusderived flavonoids, flavonoids, flavonoes comprise approximately 95 % of the total flavonoids (Peterson *et al.*, 2006). And flavones, flavonols and polymethoxylated flavonoids are present in lower concentration. In addition, some of flavonoids are unique to Citrus plants. Citrus-derived flavonoids are present in glycoside or aglycone forms, and usually do not occur naturally as aglycones but rather as glycosides, in which the aglycones are linked to a sugar moiety. Among the aglycone forms, naringenin, hesperetin, apigenin, nobiletin, tangeretin and quercetin are widely detected. For glycoside forms, D-glycosides, C-glycosides, rutinosides, glucosides and neohesperidosides are common. Naringin (neohesperidoside), neohesperidoside), narirutin (rutinoside), and hesperidin (rutinoside) are commonly present in major quantities. Sinensetin, isosinensetin, nobiletin, tangeretin, which all belong to polymethoxylated flavones, exist only as aglycones because the binding sites for sugar moieties are not occupied by hydroxyl moieties (Nielson *et al.*, 2000).

#### **Bioactivities of Citrus fruits**

Owing to these metabolites, Citrus fruits exhibit plentiful bioactivities including anti-oxidant, anti-inflammatory, anti-cancer, anti-microbial and anti-allergy activities, as well as cardiovascular effect, neuroprotective effect, hepatoprotective effect, obesity control, etc. Note that

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flavonoids (especially flavanone, flavanonol and methoxylated flavones) are more active compared to other secondary metabolites in Citrus for their remarkable various bioactivities.

Flavonoids The juices from green and ripe chinotto (*C. myrtifolia*), which were full of flavonoid, was tested by DPPH· radical bleaching and superoxide-anion scavenging, and it was shown that immature chinotto fruits, in particular, yield a juice with a remarkable anti-oxidant power. The anti-oxidant activity of the flavonoid mixture isolated from the Citrus peel was determined in terms of the DPPH· and ABTS· + scavenging and the reducing power assay in a concentration range from 25 to 500 mg/L, and its anti-oxidant activity increased in a dose-dependent manner the results of the anti-oxidant capacities and total phenolic acids contents of the Tarhana samples (Kilci and Gocmen, 2014). The anti-oxidant potency composite index showed wide variations, ranging from 58.84 to 98.89 in the 14 studied wild mandarin genotypes native to China, due to different phenolic compounds' levels, including phenolic acids.

Dgiwara *et al.* (2003) found that caffeic, chlorogenic, and ferulic acids scavenged various radicals, such as superoxide anions and hydroxy radicals. Citric acids from Citrus have been found to show anti-oxidant activity in lipopolysaccharide (LPS)-treated mice. Korani *et al.* (2014) demonstrated that gallic acid has a beneficial activity against 2VD-induced cognitive deficits via enhancement of the cerebral anti-oxidant defense. Among the phenolic acid group, gallic acid with three hydroxyl groups on the aromatic ring was the strongest anti-oxidant (Karamac *et al.*, 2005). In contrast, the monosubstitutedphenolic acids (p-coumaric acid, o-coumaric acid, and 4-OH-phenylacetic acid) showed very low activity. In addition, the radical-scavenging activities of phenolic acids are related to their hydroxyl group characteristics in the order: gallic > gentisic > syringic > caffeic > protocatechuic > sinapic > ferulic > isoferulic > vanillic > p-coumaric > o-coumaric > m-coumaric > salicylic >> p-hydroxybenzoic (Jabri Karoul and Marzouk, 2013).

### Antimicrobial action of Citrus fruits

Fruit juice of *C. limon* inhibited clinical isolates of *Staphylococcus aureus, Escherichia coli, Klebsiellaaerogenes, and Klebsiella pneumonia* (Pradeep *et al.,* 2007). Peel extracts of *C. sinensis* has marked antibacterial action while C. aurantium revealed higher antifungal activity against Colletotrichumcapsici in comparison with each other (Madhuri *et al.,* 2014). Aqueous extracts of peel and juice from fresh and dried Citrus and sweet lemon reported antimicrobial action against six Gram-positive and eight Gram-negative bacterial and one yeast isolates (Nada and Zainab, 2013).

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*C. sinensis* seed oil is highly effective as an antifungal agent against Lentinussajor-caju, which caused white rot in wood hence, it can be used as a preservative agent in the management of wood infected with white rot fungi (Yekeen *et al.*, 2014).Propionibacterium acnes, bacterial species that causes Acne vulgaris is reported very sensitive to C. limon (Shinkafi and Ndanusa, 2013). Methanolic extract of C. sinensis fruit peel was able to inhibit all the bacteria (*Bacillus subtilis, S. aureus, E. coli*) and fungi (*Candida. albicans* and *Aspergillusniger*) with different degrees of inhibition (Dhiman *et al.*, 2012). The high biological potential of essential oil of *C. limon*, against multidrug resistant *Acinetobacter spp.* was reported (Guerra *et al.*, 2013).

#### **Estrogenic activity**

Significantly decrease sperm count was found after administration of alcoholic extract and its fractions of *C. limonum* seeds for 60 days but the count returned to normal after 90 days. Sperm count reduction and atrophic changes in testis and epididymis suggest reduction in male sex hormone that is androgen as the mechanism of anti-fertility effect (Kulkarni *et al.*, 2012). The petroleum ether extract proved to retain high estrogenic activity in immature female rats (EI-Alfy *et al.*, 2012). Petroleum ether extract of *C. medica* seeds exhibited estrogenic effects in rats which include an increase in uterine weight and vaginal epithelial cell cornification. The opening of vagina on the 5th day and cornification of vaginal epithelial cells was found in 30-day-old immature rats. Alcohol and chloroform extract of *C. limonum* seeds showed reversible anti-fertility effect in mice by virtue of its anti-zygotic action. *C. medica* Linn. (peel) was used as the traditional/folkloric medicine for anti-fertility activity (Kulkarni *et al.*, 2005).

### **Description of lemon**

Lemon constitutes an important fresh fruit group even though it is not eaten fresh as mandarins and oranges. They usually have high acid content although acidless cultivars also exist (Ortiz, 2002). It is used primarily for drinks and fresh juice or lemonade, cooking and flavouring, especially in the making of lemon pies, lemon cakes, candies, jams and marmalades, and also for medicinal purposes due to its high content of vitamins. The fruit is generally oval to elliptical with characteristic necks and nipples. The peel is yellow at maturity and has prominent oil glands. The flesh is pale yellow in colour and very sour. There are three major groups of lemons: the Femminello, the Verna and the Sicilian groups (Ortiz, 2002).

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## Scientific classification of lemon

Domain: Eukaryota

Kingdom: Plantae

Phylum: Spermatophyta

Subphylum: Angiospermae

Class: Dicotyledonae

Order: Rutales

Family: Rutaceae

Genus: Citrus

Species: *Citrus limon* 

# **Materials and Methods**

Autoclave, incubator, petri dishes, test tubes, conical flask, colony counter, syringes, knife, Agar slant bottles, slides, microscope, filter paper, paper tape, bijou bottles, test tube rack, wire loop, lemon ( ripe and unripe ).

## Sample Collection and Extraction

The samples (lemon fruits) were purchased from total market Oko in Anambra state, Nigeria. None of this sample had spoilt at the time of this investigation.

## Sample preparation

10 lemon fruits (ripe and unripe) were aseptically squeezed into active sterile bijou bottles to obtain its juice.

## **Test Organisms**

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The test organisms that were used for this work were collected from Nation Agency for Food and Drug Administration and Control in Agulu. The organisms include: *Staphylococcus aureus, Bacillus subtilis. Escherichia coli and Pseudomonas aeruginosa.* 

#### Antimicrobial assay.

The antimicrobial activity was performed by agar disc diffusion method. The bacterial strains were grown in nutrient broth, Nutrient agar was the media used to study the bacteria susceptibility. The broth culture were grown 24hours and serially diluted in the same broth c sterilized at 121°C for 15minutes. The 24hours broth culture containing approximately 1.7x10°cfu/ml, 2.6x10°cfu/ml, 2.6x10°cfu/ml and 5,0x10°cfu/ml. for *Escherichia coli, Staphylococcus aureus, Bacillus subtilis and Pseudomonas aeruginosa*. respectively as determined by plate count method sterile swab stick was used.

## Minimal Inhibitory Concentration (MIC)

The extract that gave up to 30mm zone of inhibition were used to determine the MIC on the respective organisms they inhibited (Staphylococcus aureus and Pseudomonas auriginosa)

The extracts were diluted with sterile distilled water to give the following concentrations (v/v) 50%, 30%, 20%, 15%, 10%, and 5%, corresponding to 2.5ml, 1.5ml, 1ml, 0.75ml, 0.5ml and 0.25ml of the extracts in 5ml of broth. These were inoculated with 0.5ml of 10<sup>-3</sup> dilution of 24hours broth culture of the test organisms; *Staphylococcus aureus* and *Pseudomonas auriginosa*. Tubes without extract and that without culture were also prepared to serve as positive and negative control respectively. These were prepared for 24hours at 37°C and the MIC determined as the lowest concentration with no visible growth.

#### Minimum Bactericidal Concentration (MBC)

Test tube of culture showing no visible growth were gently streaked on NA plates and incubated for 24hours to determine the MBC which is the lowest concentration of the extract that did not allow growth in NA after 24hours incubation.

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## RESULTS

The result of the antimicrobial activity test showed that the juice extracts (riped and unriped)lemon, were effective to some of the organisms such as; *Staphylococcus aureus* and *Pseudomonas aeruginosa* as shown in table1and plates 1-5.

## **Minimal Inhibitory Concentration (MIC)**

The MIC is the lowest concentration that did not show visible growth in dilution tube. The result of the MIC is as shown in table2.

## Minimum Bactericidal Concentration (MBC)

The MBC is the concentration of the extracts that did not allow the growth of the organism after 24hour incubation on Agar. The result of

the MBC is as shown in table 3.

## Table 1: Antimicrobial activity of the juice extract against some bacteria strains tested.

Test Organisms	Control	Zone of inhibition Ripe lemon Unrip				
S. aureus	30	30	24			
Pseud. aeruginosa	7. 33	28	30			
Bacillus subtilis	38	20	Nil			
E. coli	25	Nil	23			

Table 2: The result for the MIC of the juice extracts on <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i>					
Extracts	Test Organisms	MIC			
Ripe Lemon	Staphylococcus aureus	30% v/v			
Unripe Lemon	Pseudomonas auriginosa	30% v/v			

Table 3: The result for th Extracts	e MBC of the juice extracts on Test Organisms	Pseudomonas	<i>s spp</i> . and <i>Staphylococcus aureus</i> MBC	
Ripe Lemon	Staphylococcus aureus	30% v/v		
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Unripe Lemon

Pseudomonas auriginosa

30% v/v

## DISCUSSION

This study was carried out to investigate the possible antimicrobial activity of lemon (*Citrus limon)*on selected microorganisms (*Pseudomonas aeruginosa, Staphylococcus aureus, Escherichia coli, Bacillus subtilis*.

The result of the test showed antimicrobial activities of ripe lemon juice on *Staphylococcus aureus* (30mm), *Pseudomonas aeruginosa* (28mm), *Bacillus subtilis* (20mm) and no zone of inhibition in *Escherichia coli, w*hile in unripe lemon juice showed S*taphylococcus aureus* (24mm), *Pseudomonas aeruginosa*. (30mm), *Escherichia coli* (23) and no zone of inhibition in *Bacillus subtilis*.

This result partially agrees with the finding of Ammara *et al.* (2009) on the zone of inhibition showed on *Staphylococcus aureus* (30mm) but varied with the inhibition showed on *Escherichia coli* (8mm). *Pseudomonas spp* (16mm), *Bacillus spp* (20mm)

This difference could be as a result of specie of lemon fruits used, the laboratory environment, or the method of extraction of the juice. The antimicrobial effect of these fruits could be attributed to the phytochemical constituents present in it, which are known to confer certain health such as antimicrobials.

The result of this work has shown that lemon juice (ripe and unripe) can be used in the treatment of urinary tract infections, diarrhea, wound, virginal discharge, cholera and dysentery.

## CONCLUSION

This study shows that lemon ( ripe and unripe) juice contains valuable antimicrobials. It has been investigated to inhibit the growth of *Staphylococcus aureus, Pseudomonas, Bacillus* and *Escherichia coli*. Therefore lemon can provide treatment and remedies against human infection caused by these organisms.

## RECOMMENDATION

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In view of the above results, the natural and traditional use of lemon and lime (ripe and unripe) should be continued, because it contains

many antimicrobials. It is also recommended for human consumption, pharmacology and herbal medicines for the treatment of bacterial

infections and diseases.

## References

Abeysinghe, D. E. (2010). Isolation, characterization and antibacterial activity of Orange fruits. Journal of Microbiology 4(6):7-15.

Allison, S.D., Jennifer, and B.H.M. (2008). Resistance, resilience, and redundancy in microbial communities. PNAS. 105(suppl. 1): 11512-11519.

- AL-Waili, N.S., Akmal, M., AL-Waili, F.S., Saloom, K.Y., and Ali, A. (2005). The antimicrobial potential of *Citrus species* on some microbial isolates. Med Sci Monit.11: 433-438.
- Amal, S.D. (2014). Antibacterial Activity of Bee and Yemeni Sidr Honey Against Some Pathogenic Bacterial Species. *Int. J. Curr. Microbiol. App. Sci.*, 3(10): 1015-1025.
- Ammara, H, Sarker, V., and Nahar, C. (2009). Antimicrobial Activities of *Staphylococcus aureus, Salmonella spp, Pseudomonas auriginosa, Eschericia coli*, and *Bacillus spp* Drange fruits; 4(1): 63-78.
- Aranganathan, S, and Nalini, N. (2009). Efficacy of the potential chemopreventive agent, hesperetin (*Citrus flavanon*e), on 1, 2-dimethylhydrazine induced colon carcinogenesis. Food Chem Toxicol 47:2594–2600
- Aruoma, D. I, Landes, B, Ramful-Baboolall, D, Bourdon, E, Neergheen-Bhujun, V, Wagner, K. H, and Bahorun, T. (2012) Functional benefits of citrus fruits in the management of diabetes. Prev Med 54:S12–S16
- Ball, D. (2007). The chemical composition of honey. J. Chem. Educ., 84: 1643-1646.
- Barchiesi, F., Arnaldo, L.C., Deanna, A.M., and Michael, G.R. (2004). Comparative Study of Broth Macrodilution and Microdilution. Techniques for In Vitro Antifungal Susceptibility Testing of Yeasts by Using the National Committee for Clinical Laboratory Standards' Proposed Standard. Journal of Clinical Microbiology, 32(10): 2494- 2500.

Bhavsar, S, Joshi, P, Shah, M, and Santani, D. (2007). Investigation into hepatoprotective activity of Citrus limon. Pharm Biol;45:303-11.

Bogdanov, S., Jurendic, T., Sieber, R., and Gallmann, P. (2008). Honey for nutrition and health: A review. J Am CollNutr. 27:677-689.

- Bui, F. U. (2010). Evaluation of the chemical composition of lime exudates used in herbal medicine in South Eastern Nigeria. *Afr. J. Trad.* Comp. Alt. Med. 5(2):194-200.
- Burt, S.A. (2004). "Essential oils: Their antibacterial properties and potential applications in foods: A review". *International Journal of Food Microbiology*, 94(3): 223-253.
- Canter, A.A, and Ernst, J. K. (2004). The phytochemical analysis and antibacterial effects of lime and lemon. Int J Herb Pharmacol Res, 4(2):10-6.
- Caristi, C, Bellocco, E, Panzera, V, and Toscano, G. (2003). Flavonoids detection by HPLC-DAD-MS-MS in lemon juices from Sicilian cultivars. J Agric Food Chem 51:3528–3534
- Casimiro, M, Margarita, G, Danice, R, and Judilynn, N. (2010). Evaluation of the hepatoprotective activity of Citrus microcarpa Bunge (Family Rutaceae) fruit peel against acetaminophen-induced liver damage in male BFAD- Sprague Dawley rats. *International Journal of Chemical Environmental Engineering*,1:127-32.
- Cheesbrough, M. (2006). District Laboratory Practice in Tropical Countries (2nd edn). Cambridge University Press, New York; 178-194.
- Chen, J, Montanari, A. M, and Widmer, W. W. (2017). Two new polymethoxylated flavones, a class of compounds with potential anticancer activity, isolated from cold pressed dancy tangerine peel oil solids. *J Agric Food Chem* 45:364–368
- Committee, N. P. (2010). Pharmacopoeia of People's Republic of China, vol 2. China Medicinal Science and Technology Press, Beijing
- Corbo, M, Kokate, G, Amin, H. A. S., Hanna, A. G., and Mohamed, S. S. (2008). Comparative studies of acidic and enzymatic hydrolysis for saponin. Biocatalysis and Biotransformation, 29(6), 311–319.
- Davidson, P.M., and Naidu, A.S. (2000). Phytophenols. In A. S. Naidu (Ed.), Natural food antimicrobial systems. Boca Raton, FL: CRC Press; pp. 265-294.
- Demonty, I, Lin, Y, Zebregs, Y. E, and Vermeer, M. A. (2010). The citrus flavonoids hesperidin and naringin do not affect serum cholesterol in moderately hypercholesterolemic men and women. J Nutr 140:1615–1620.

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## SCIENCE RESEARCH

# FEDERAL POLYTECHNIC OKO, ANAMBRA STATE

- Denise, M., Dubet, Silva M., and Paulo, N.N. (2002). "A High Grassland Bee Community in Southern Brazil: Survey and Annotated checklist (Insecta: Apidae)". Journal of Kansas Entomological Society, 85(4): 295-308.
- Dhiman, A, Nanda, A, Ahmad, S, and Narasimhan, B. (2012). In vitro antimicrobial status of methanolic extract of *Citrus sinensis* Linn.fruit peel. Chron Young Sci;3:204-8.
- Economos, C, and Clay, W. D. (2019) Nutritional and health benefits of citrus fruits. Food Nutr Agric 24:11-18
- EI-Alfy, T, Hetta, M, Yassin, N, Rahman, R, and Kadry, E. (2012). Estrogenic activity of Citrus medica L. leaves growing in Egypt. *J Appl Pharm Sci*.2:180-5.
- García-Salas, P, Gómez-Caravaca, A. M, and Arráez-Román, D. (2013). Influence of technological processes on phenolic compounds, organic acids, furanic derivatives, and antioxidant activity of whole lemon powder. Food Chem 141:869–878
- Giuseppe, H, Kinyanjui, T, Gitu, P. M, and Kamau, G. N. (2007). Potential antitermite compounds from *Juniperus procera* extracts. Chemosphere 41(7):1071-1074.
- Guerra, F, Mendes, J, Oliveira, W, Souza, F, Trajano, V, and Coutinho, H. (2013). Antibacterial activity of the essential oil of Citrus limon against multidrug resistant Acinetobacter strains. Rev Bras Farm;94:142-7.
- He, D, Shan, Y, Wu, Y, Liu, G, Chen, B, and Yao, S. (2011). Simultaneous determination of flavanones, hydroxycinnamic acids and alkaloids in citrus fruits by HPLC-DAD-ESI/MS. Food Chem 127:880-885
- Jabri Karoui, I, and Marzouk, B. (2013). Characterization of bioactive compounds in Tunisian bitter orange (*Citrus aurantium* L.) peel and juice and determination of their antioxidant activities. Biomed Res Int 2013: 345415
- Johann, Y, Philip, C and Kar, R. (2007). Antimicrobial Activity of Five Herbal Extracts Against Multi Drug Resistant (MDR) Strains of Bacteria and Fungus of Clinical Origin. Molecules; 14: 586-597.
- Kalpa, G, Khan, A. V, Khan, A. A, and Shukla, I. (2012). In vitro Antibacterial Potential of Melia azedarach Crude Leaf Extracts Against Some Human Pathogenic Bacterial Strains. Ethnobotanical Leaflets 12:439-445.
- Karamac, M, Kosinska, A, and Pegg, R.B. (2005). Comparison of radical-scavenging activities for selected phenolic acids. *Polish J Food Nutri Sci* 14:165–170
- Kato, Y., Natsuki, U., Asuna, M., Daiki, M., and Noritoshi, K. (2012). "Identification of a novel glycoside, leptosin, as a chemical marker of manuka honey". Journal of Agricultural and Food Chemistry, 60(12): 3418-23.
- Kawaii, S, Tomono, Y, Katase, E, Ogawa, K, and Yano, M. (2019). Quantitation of flavonoid constituents in citrus fruits. J Agric Food Chem 47:3565–3571
- Kawaii, S., Satoru, K., Yasuhiko, T., Eriko, K., Kazunori, D. and Masamichi, Y. (2000). "Quantitative study of flavonoids in leaves of Citrus plants". *Journal of Agricultural and Food Chemistry*, 48(9): 3865-3871.
- Keizo, Y., Yoichi, H., Intetsu, K., Masanari, I., and Akira, D. (2012). A report from the committee on microbroth dilution antimicrobial susceptibility testing Issues in antimicrobial susceptibility testing of *Haemophilus influenza*. a comparison of the Japanese Society of Chemotherapy lysed horse blood and the CLSI HTM broth microdilution methods. *J Infect Chemother*. 18:134-143.
- Kelebek, H, and Selli, S. (2011). Determination of volatile, phenolic, organic acid and sugar components in a Turkish cv. Dortyol (Citrus sinensis L. Osbeck) orange juice. *J Sci Food Agric* 91:1855–1862
- Keles, F, Chris, P and Jonathan, N. (2001). Phytochemical Analysis and Antimicrobial Activity of Scoporiaduals and NympheaCotus. *Autrallian Journal of Basis Applied Sciences* 3 (4): 3975-3979.
- Kilci, A, and Gocmen, D. (2014). Phenolic acid composition, antioxidant activity and phenolic content of tarhana supplemented with oat flour. Food Chem 151:547–553
- Koc, A.N., Silici, S., Ercal, B.D., Kasap, F., Hörmet-Öz, H.T., Mavus- and Buldu, H. (2008). Antifungal activity of Turkish honey against Candida spp. and Trichosporonspp: An in vitro evaluation. Med Mycol. 47:707-12.
- Korani, M. S., Farbood, Y., Sarkaki, A., Fathi, Moghaddam, H., Taghi, and Mansouri, M. (2014). Protective effects of gallic acid against chronic cerebral hypoperfusion-induced cognitive deficit and brain oxidative damage in rats. *Eur J Pharmacol* 733:62–67
- Kulkarni, T, Mateenuddin, M, Bodhankar, S, and Saharabudhe, R. (2012). Reversible anti- fertility effect of lemon seeds (*Citrus limonum*) in Male Albino Rats. IJRPBS;3:545-50.
- Kulkarni, T. R, Kothekar, M. A, and Mateenuddin, M. (2005). Study of antifertility effect of lemon seeds (*Citrus limonum*) in female albino mice. *Indian J Physiol Pharmacol*,49:305-12.
- Kwakman, P.H., Van den Akker, J.P., Güçlü, A., Aslami, H., and Binnekade, J.M. (20080. Medical-grade honey kills antibiotic resistant bacteria in vitro and eradicates skin colonization. Clin Infect Dis. 46 (11): 1677-16682.
- Li, M.M. (2000). Handbook of Africa Medicinal Plants CRC Press, Inc. London p. 19.

SCIENCE RESEARCH

# FEDERAL POLYTECHNIC OKO, ANAMBRA STATE

- Liu, Y, Liu, Z, Wang, C, Zha, Q, Lu, C, Song, Z, Ning, Z, Zhao, S, Lu, X, and Lu, A. (2014). Study on essential oils from four species of Zhishi with gas chromatography-mass spectrometry. *Chem Cent J* 8:1–8
- Liu, Z, Liu, Y, Wang, C, Song, Z, and Zha, Q. (2012). Discrimination of Zhishi from different species using rapid-resolution liquid chromatography-diode array detection/ultraviolet (RRLC-DAD/UV) coupled with multivariate statistical analysis. *J Med Plants Res* 6:866–875
- Madhuri, S, Hegde, A, Srilakshmi, N, and Prashitha, K. (2014). Antimicrobial activity of *Citrus sinensis* and *Citrus aurantium* peel extracts. JPSI;3:366-8.
- Mandal, S. (2011). "Honey: its medicinal property and antibacterial activity". Asian Pacific Journal of Tropical Biomedicine, 1(2): 154-160.
- Manisha, D.M., and Shyamapada, M. (2011). Its medicinal property and antibacterial activity. Asian Pacif. J. Trop. Biomed. 1(2): 154-160.
- Maurya, R, Srivastava, S, Kulshreshta, D. K, and Gupta, C. M. (2004). Traditional remedies for fertility regulation. Curr Med Chem;11: 1431-50.
- Mavric, E., Wittmann, S., Barth, G., and Henle, T. (2008). Identification and quantification of methylglyoxal as the dominant antibacterial constituent of manuka (*Leptospermum scoparium*) honeys from New Zealand. Molecular Nutrition and Foods Research, 52:483-489.
- Mayer, B.H., and Williams, L. (2004). Women's health: A guide to health promotion and disorder management. Lippincott Williams and Wilkins; Philadelphia; p. 405.
- Molan, P.C. (2012). "The Antibacterial Activity of orange fruits. Variation in potency of the antibacterial activity". 73: 59-76.
- Morgan, R.H. (2002). *The Alkaloids: Chemistry and Physiology* Vol. viii. New York. Academic Press p. 673.
- Mshelia, B.M., Adeshina, G.D., and Onaolapo, J.A. (2017). The Antibacterial Activity of lime and Lemon Juice against *Streptococcus pregenes* Isolates from Respiratory Tract Infections. Adv Biotech & Micro. 4(5): 001-008.
- Nada, K, and Zainab, A. (2013). Antimicrobial activity of different aqueous lemon extracts. J Appl Pharm Sci,3:74-8.
- Nascimento, G. G. F. Locatelli, J. Freitas, P. C. and Silva, G. L., (2000). Antibacterial Activity of Plant Extracts and Phytochemicals on AntibioticResistant Bacteria. *Brazilian Journal of Microbiology*, 31 (4): 247-256.
- Negi, S, and Anand, B. (2010). Analgesic activity of fruit decoction of Citrus medica Linn. Journal Pharmacy Resource, 3:2119.
- Nielsen, S, Breinholt, V, Cornett, C, and Dragsted, L. (2000). Biotransformation of the citrus flavone tangeretin in rats. Identification of metabolites with intact flavane nucleus. Food Chem Toxicol 38:739–746
- NPCS. (2012). "Handbook on Agro Based Industries (2nd Edition)". Published by Niir Project Consultancy Services, Delhi (India), 75-78.
- Ogiwara, T, Satoh, K, Negoro, T, Okayasu H, Sakagami, H, and Fujisawa, S. (2003). Inhibition of NO production by activated macrophages by phenolcarboxylic acid monomers and polymers with radical scavenging activity. Anticancer Res 23:1317–1323
- Ortiz, F. C. (2002). Characterization of wild-type *Salmonella spp* and their susceptibility to "Mist Enterica", An Anti-Typhoid Herbal Preparation. PhD. Thesis, University of Ghana.
- Ortuño, J. M, Kenji, G. M, and Gachanja, A. N (2006) Anti-termite and antimicrobial properties of paint made from Thevetia peruviana (Pers.) Schum. oil extract. *Afr. J. Pharm.* Pharmacol. 4(2):87-89.
- Pal, B, Mehmood, B, Dar, C and Ghous, T. (2003) short communication: in vitro assessment of antioxidant, antibacterial and phytochemical analysis of plant seeds. *Pak J. pharm Sci.* 2015: 28:231-9.
- Patil, S. J. and Patil, S. B. (2015). Estrogenic activity of petroleum ether extract of seeds of *Citrus medica* on immature albino rats. *Int J Green Pharm* 2008;2:91-4.
- Paulus, H.S., Kwakman, Johannes, P.C., Van-den, A., and Ahmet, G. (2008). Medical-grade antibiotic resistant bacteria in vitro and eradicates skin colonization. Clinical Infectious Diseases, 46: 1677-82.
- Pawinee, P, Thirayudh, G, and Aporn, C. (2019). Antifertility effect of *Citrus hytrix. J Ethnopharmacol*,13:105-10.
- Percy, D. W, Adcock, J. L, Conlan, X. A, Barnett, N. W, Gange, M. E, Noonan, L. K, Henderson, L. C, and Francis, P. S. (2010). Determination of *Citrus aurantium* protoalkaloids using HPLC with acidic potassium permanganate chemiluminescence detection. Talanta 80:2191–2195.
- Peterson, J. J., Beecher, G. R., Bhagwat, S. A., Dwyer, J. T., Gebhardt, S. E., Haytowitz, D. B., and Holden, J. M. (2006). Flavanones in grapefruit, lemons, and limes: a compilation and review of the data from the analytical literature. *J Food Compost Anal* 19:S74–S80
- Pradeep, M, Wadher, T, and Gomashe, A. (2007). Antibacterial activity of *Citrus limon* fruit juice against clinical isolates of human pathogens. *Asian J Microbiol Biotechnol Environ Sci*,9:129-32.
- Ramalivhana, J. N, Obi, C. L, and Samie, A. (2014). Antibacterial activity of honey and medicinal plant extracts against Gram negative microorganisms. *Afr. J. Biotechnol.*, 13(4): 616-625.
- Ramon-Laca, V. C. (2003). African tropical plant gums: grossly unexploited carriers or adjuncts in drug delivery systems. Extraction; 3:4.
- Ritcher, M. A. (2003). Depletion of forest resources in South Eastern Nigeria. The Environmentalist; 21(3):197–203.

## INTERNATIONAL JOURNAL APPLIED SCIENCE RESEARCH, INJASR. VOL. 1, JUNE 2021

# SCIENCE RESEARCH

# FEDERAL POLYTECHNIC OKO, ANAMBRA STATE

- Schmidt, M. E, and Wider, R. H (2005). Phytochemical content and antioxidant activity of six diverse varieties of whole wheat. Food Chem.119(1):249-257.
- Shayeste, B.B., Gholam, R.M., Batoul, P., and Majid, V. (2013). Comparison of the effect of honey and miconazole against Candida albicans in vitro. Adv Biomed Res. 2: 57.
- Shinkafi, S, and Ndanusa, H. (2013). Antibacterial activity of Citrus limon on Acne vulgaris (pimples). Int J Sci Invent Today, 2:397-409.
- Sivakumar, N, and Venkataraman, R. (2010). Phytochemical and pharmacological studies on plant waste materials. Pharm Sin;1:1-6.
- Soetan, K.O., Dyekunle, M.A., Aiyelaagbe, D.O., and Fafunso, M.A. (2006). Evaluation of the antimicrobial activity of saponins extract of Sorghum Bicolor L. moench. *African Journal of Biotechnology*, 5(23): 2405-2407.
- Sohn, H.Y., Son, K.H., Kwon, C.S., and Kwon, G.S. (2004). Antimicrobial and cytotoxic activity of 18 prenylated flavonoids isolated from medicinal plants: Morusalba L., *Morus mongolica* Schneider, *Broussnetia papyrifera* (L.) Vent, *Sophora flavescens* Ait and *Echinosophora koreensis* Nakai. Phytomedicine, 11(7-8), 666-672.
- Somnath, D., Nilanjana, M., Susmita, B., Puja, G., Snighadha, M., Moumi, D., and Siddhartha, C. (2017). Investigate the Antimicrobial Activity of Raw Lemon and Honey against Human Enteric Pathogens in vitro. *International Journal of Ayurvedic and Herbal Medicine*, 7(1): 2469-2474.
- Sood, S. Arora, B. Bansal, S. Muthuraman, A. Gill, N. S. and Arora, R. (2009). Antioxidant, anti-inflammatory and analgesic potential of the *Citrus decumana* L. peel extract. Inflammopharmacology;17:267-74.
- Sood, S, Bansal, S, Muthuraman, A, Gill, N, and Bali, M. (2009). Therapeutic potential of Citrus medica L. peel extract in carrageenan induced inflammatory pain in rats. *Res J Med Plant*, 3:123-33.
- Stickel, Y, Mucha, G, Kyriazakis I. and Whittenmore C.T. (2005). Whittenmore's Science and Practice of Pig Production. Oxford: Wiley Black well.
- Stohs, S. J, Preuss, H. G, and Shara, M. (2011). The safety of *Citrus aurantium* (bitter orange) and its primary protoalkaloid p-synephrine. Phytother Res PTR 25:1421–1428
- Sun, Y, Qiao, L, Shen, Y, Jiang, P, Chen, J, and Ye, X. (2013). Phytochemical profile and antioxidant activity of physiological drop of citrus fruits. *J Food* Sci 78:C37–C42.
- Swingle, W. T. (2017). The botany of Citrus and its wide relatives, vol 1. University of California Press, Berkeley
- Tanaka, T. (2017). Fundamental discussion of Citrus classification. Stud Citrol 14:1–6 11. Barret H, Rhodes A (1976) A numerical taxonomic study of affinity relationships in cultivated Citrus and its close relatives. Syst Bot 1:105–136
- Teke, G.N., and Betie, E.N. (2016). Antibacterial activity of three Cameroonian honey types against some pathogenic species. *Int. J. Biol. Chem. Sci.* 10(6): 2477- 2484.
- Tilburt, J, Lewis, W.H. and Manony, P.F.E (2008). Medical Botany: Plants Affecting Man's Health. John Wiley and Sons. New York, USA. P. 240.
- Van, C, Terry, P and Okwu, D. E. (2007). Investigation into the Medicinal and Antibacterial Effect of Orange. Baker Ph. D Thesis Michael Okpara University of Agriculture Umudike, Nigeria pp. 20-31.
- Vineetha, N., Vignesh, RA., and Sridhar, D. (2015). Preparation, Standardization of Antibiotic Disc and Study of Resistance Pattern for First-line Antibiotics in isolates from clinical samples. *International journal of applied research*, 1(11): 624-631.
- Yekeen, M, Ajala, D, and Alarape, A. (2014). Antifungal activities of *Citrus sinensis* seed oil against Lentinussajor-caju. Adv Appl Sci Res;5:109-13.
- Young, M. (2012). Antioxidant, antibacterial, antifungal activities and phytochemical analysis of lime and lemon leaves extracts. J. Med. Plant Res. 7(6):243-249.